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Hair Growth Stimulation by a Natural Remedy: Animal Studies

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Abstract

Hair follicles normally regenerate cyclically and this makes them a popular model for investigations of regeneration and aging. We studied the effects on mouse hair follicles of the Satura® Rosta remedy manufactured from plant extracts and used to stimulate hair growth restoration. A prolonged application of the remedy to the skin of various strains of mice and of outbred white mice did not induce detectable negative effects and caused a significant increase of mean hair follicle size and hair root volume as well as skin thickening due to adipose elaboration. The volume increase took place mainly among the largest hair follicles. Mouse skin thickness is known to fluctuate cyclically in coordination with the waves of hair growth; anagen regions are thicker due to adipose increase. Satura® Rosta did not abrogate hair growth waves. Apparently the observed effects could be explained by the remedy-induced prolongation of the anagen phase. Although the effect of Satura® Rosta on fur appearance was insignificant in young animals, experiments with old mice exhibiting alopecia resulted in hair regrowth in alopecia regions. Transcriptome analysis of Satura® Rosta action on dermal fibroblasts *in vitro* revealed significant changes of the expression of genes associated with the cell cycle, developmental processes and lipid metabolism.

Keywords: Hair Growth; Alopecia; Mice; Hair Follicles; Anagen; "Satura® Rosta"; Adipose; Aging; Transcriptome.

Introduction

The creation of a remedy for effective hair growth stimulation has for many years been a dream of cosmetologists through the entire world. Although alopecia and hair loss are treated by cosmetologists, they are also generally accepted marks of aging. In recent years, there has been a growing interest to hair follicle studies because hair follicle can serve as an informative model for investigation of cell senescence, aging of tissues, organs and the whole body.

Hairs grow cyclically. Several periods can be distinguished within the hair growth cycle. The growing phase of a hair follicle is called anagen, after it occurs catagen-the period of degeneration, and at the end telogen-the rest phase [1]. In human scalp the hair growth cycle spans up to 7 years, in mouse skin-a few weeks. Hair follicles are capable to return to growth after a prolong period of rest.

There is a great intensity of metabolism in growing follicles. The beginning of the growth is prepared by the accumulation of building materials and energy including the development of subcutis—a thick layer of skin located under the derma and composed mainly of adipocytes. During anagen, the follicles are completely immersed into this adipose layer and enhanced growth of blood capillaries surrounding follicles is observed. In contrast to humans, most of the mice hairs grow synchronously. There are waves of hair growth moving along the body. Waves are strictly oriented in young animals, later the waves became weaker and finally islands of hair growth appear instead of waves. Growth waves cyclically alter skin thickness by influencing adipose elaboration and regression [2]. Skin aging is associated with adipose reduction, decreased cellularity and reduction of metabolism [3]. Age-related skin deterioration should influence hair growth; however hair greying and alopecia develop earlier than classical phenotype of old skin.

There are many types of alopecia that differ in underlying mechanisms and the degree of development. Among etiological factors of alopecia are heredity, autoimmune processes, aging or premature aging, consequences of anti-cancer therapy and even psychological stress. Pathogenesis of different kinds of alopecia is not completely understood therefore the market offers an impressive number of different medications of which the majorities are ineffective or even have negative side effects.

The most widespread kind of alopecia is the androgenetic one (AGA). It constitutes around 90% of cases. It is characterized by the appearance of areas of baldness in frontal and parietal zones of man scalp and hair rarefaction in the same areas in women. Hair cycles are shortened as a result of diminished anagen that leads to the vellus transformation of scalp hairs, hair follicle miniaturization, and a decrease of hair size up to their complete disappearance [4]. All the hairs in affected areas can be involved in this process. Along with hair miniaturization, the production of pigment ceases. AGA is a very widespread problem: about 50% of men beyond the age of 50 years and 50% women beyond the age of 60 years are subject to AGA [5]. Some affected individuals experience great psychoemotional stress, often leading to a reduction of quality of life.

From the early 1940s it has been known that the process of AGA development is mediated mainly by androgen [6]. Disease is characterized by the increased responsiveness of dermal papilla in specific locations to testosterone that leads them to increased secretion of cytokines such as TGF- β and DKK-1. TGF- β [7-9] is known as an anagen inhibitor in the hair cycle. DKK-1 [10] inhibits the growth of the outer root sheath cells and induces apoptotic cell death. It is likely that these mediators determine an androgen induced inhibition of hair growth and early catagen induction in AGA [11].

There are two US Food and Drug Administration-approved therapies for treatment of AGA - Minoxidil and Finasteride. Minoxidil is a potassium channel opener, initially used for hypertension treatment. Minoxidil appears to increase the duration of the anagen phase, and its angiogenic effects result in the maintenance of hair follicles sizes [12-14].

Finasteride initially introduced for treatment of benign prostatic hyperplasia selectively inhibits the type 2 5 α -reductase isoenzyme responsible for converting testosterone to dihydrotestosterone (DHT), the putative hormonal modulator of AGA in men. Finasteride reduces serum and scalp DHT concentrations by approximately 50%. Finasteride may inhibit or reverse miniaturization of hair follicles. Finasteride is generally well tolerated during a long-term daily application [15-17]. Controversial data exist regarding the therapeutic efficacy of finasteride in treating females AGA [18-20].

Although Minoxidil and Finasteride both retard or stop hair loss and stimulate some hair regrowth, neither medication restores all lost hair or reverses complete baldness. The existing baldness therapies were discovered by accident and precise mechanism of their action are unclear. Hypothetically there are many targets upstream and downstream of androgenic signaling pathway that can be explored for baldness treatment. Other approaches include anti-aging therapies and stimulation of tissue regeneration.

We presume that any anti-aging treatment should have hair stimulating activity. For many years, the balsam "Satura® Rosta" (RSHG) has been known as an effective remedy for baldness treatment. We began to investigate its action and discovered that RSHG can induce adipogenic differentiation [21] and activate autophagy [22]. A few years ago, we began experiments involving long-term application of RSHG to mice. The results of this work are presented here.

Materials and Methods

Remedy for stimulation of hair growth (RSHG)

Balsam "Satura® Rosta" (Manufactured by GZ Laboratories (UK) Ltd) is a preparation for baldness treatment. It has been used since 1989 in various countries (USA, Great Britain, Israel, and Russia). This preparation has been tested many times in different laboratories to comply with the safety requirements and sanitary regulations for perfumes and cosmetics. No microorganisms are revealed. Toxic properties comply with the standards and it exhibits no skin irritant or allergenic effects [21]. The exact composition (Intellectual Property and Methodology) of "Satura® Rosta" is wholly owned and trade marked by GZ Laboratories (UK) Ltd. The General Director of GZ Laboratories (UK) Ltd Dr. G.V. Zigmond has advised the authors that the basis of Satura is a processed mixture of numerous terrestrial plant and seaweed extracts.

We used balsam "Satura® Rosta" for testing on animals. For experiments with cell culture we used the basis of the balsam that constitutes only 5% of the balsam. This basis was provided by Dr. G.V. Zigmond. We prepared a 1% ethyl alcohol solution and used 0,001% final concentration.

NMR analysis

¹H NMR spectrum was recorded on an AMX III-400 spectrometer (Bruker BioSpin GmbH, Karlsruhe, Germany) with the working frequency of 400 MHz using DMSO-*d*₆ as the solvent.

Animal Studies

The mice were housed in animal quarters of the Gamalei Institute of Epidemiology and Microbiology and received chow *ad libitum*. All animals were handled in compliance with the Guide for the Care and Use of Laboratory Animals approved by the Institutional Animal Care and Use Committee at the Gamalei Institute of Epidemiology and Microbiology. Mice (Balb/c, CBA, C57Bl/6 and outbred white laboratory mice) 2-3 months old were purchased from the Stolbovaya mice hatchery. Initially every experiment included 10 control and 10 experimental mice. Mice were treated with "Satura® Rosta" every second day throughout the experiments. The balsam (2 drops ≈ 100 μl) was applied on the back between the shoulders and then spread. The animals were sacrificed at different time points after the beginning of treatment (see Results) and used for immunohistochemistry studies. The obtained skin samples (sites of "Satura® Rosta" applications) were fixed with formalin, embedded in paraffin, and the sections were prepared and stained with hematoxylin-eosin. Plucked hairs were collected at different times (see Results) from the area of balsam application. Hairs were plucked by a single movement to obtain all of them from the same site. The photography was carried out using a Nikon Diavert inverted phase-contrast photomicroscope equipped with a Nikon D5000 camera.

Cell culture

Primary cultures of human skin fibroblasts (strain 1608) were kindly provided by S.M. Terekhov (Medical Genetic Research Center, Moscow, Russia). Cells were grown in DMEM medium (PanEco, Russia) supplemented with 10% fetal bovine serum (FBS) (HyClone, USA), 2mM L-glutamine (PanEco, Russia) and 40 μg/ml gentamycin (PanEco, Russia) at 37°C in the presence of 5% CO₂. Cells were treated by the 1% ethyl alcohol solution of the basis of balsam during 24 h. Final concentration of balsam basis was 0,001%. Control cells were treated with solvent only.

RNA isolation, preparation of mRNA libraries and sequencing

All RNA procedures were described previously [23]. Briefly: samples of cells (20–30 μg) frozen in liquid nitrogen were homogenized manually. RNA was isolated using a ZR RNA MiniPrep kit. The RNA amount was estimated using a NanoDrop 1000 spectrophotometer (Nano drop, USA) and a Qubit 2.0 fluorimeter (Invitrogen, USA); RNA quality was checked using an Agilent 2100 bioanalyzer (Agilent Technologies, USA). The RNA integrity number (RIN) was no less than 7 for RNA preparations.

To prepare an mRNA library we used 3.5 μg of total RNA isolated as above. Samples were prepared for sequencing with a TruSeqRNA Sample Prep kit v. 2. A total RNA sample (3.5 μg) was diluted with 50 μL of deionized water. Poly (A) mRNA was isolated with magnetic RNA purification beads (Illumina, USA) coated with poly (T). Fragmentation of purified mRNA was also performed at this step. The first cDNA strand was synthesized on the RNA template with Super Script II reverse transcriptase (Invitrogen, United States) and specific primers. The second strand was synthesized using enzymes and primers from the same kit. The resulting dsDNA was

purified using AMPure XPBeads (Bekman Coulter, USA). Blunt ends were generated using 3'- and 5'-exonucleases. A single adenine was added to the blunt 3' ends to prevent their ligation with each other during ligation of adapters. Ligation to thymine-containing adapters was carried out, and the DNA samples were prepared for hybridization to a flowcell. PCR was run as recommended by the kit manufacturer.

The libraries were tested for quality on an Agilent 2100 analyzer; the concentration was estimated via real-time PCR on a 7500 real-time PCR system (Applied Biosystems, USA). DNA clusters were generated using a cBot device according to an Illumina protocol. The flowcell with DNA clusters obtained using cBot was used for sequencing on an Illumina HiSeq 2000 whole-genome sequencer.

Transcriptome analysis

A functional analysis of the data was performed using MetaCore software and data-base (GeneGO, Thomson Reuters, and USA). Differential expression was assumed for a gene if its expression was changed (increased or decreased) more than twice compared with the normal level (differential expression threshold $|\log_2(\text{fold-change})| > 1$). A MetaCore functional ontology analysis yielded the MetaCore Maps, MetaCore Networks, and GO Processes that were enriched in differentially expressed genes to the highest extent. To estimate the enrichment significance, *p-value* was calculated using a hypergeometric distribution equation. Biological processes with the lowest *p-values* were considered the most significant. For convenience, we used the negative logarithm (base 10) of the *p-value* ($[-\log(p\text{-value})]$).

Results

NMR analysis of RSHG

Multiplet at 3.28 - 3.46 ppm corresponds to signals of glycerol (Fig. 1). The remaining signals indicate the presence of fatty unsaturated acids. Multiplet at 5.30 - 5.32 ppm corresponds to signals CH = CH of oleic, linoleic and linolenic acids. The triplet at 2.73 ppm is characteristic for the methylene group = -CH₂- = of linoleic and linolenic acids. Thus, we see that the basis of RSHG is formed mainly by fatty acid derivatives of glycerol, in which polyunsaturated fatty acids are present.

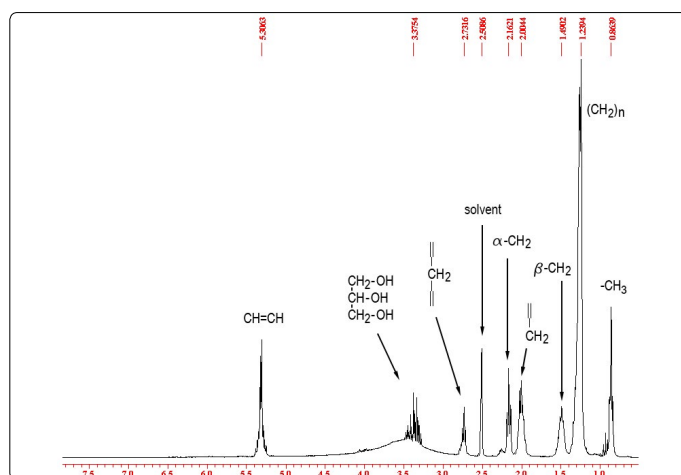


Figure 1. NMR analysis of RSHG

Long term RSHG application increases hair follicle size

A series of experiments with continuous RSHG application to the skin was performed. Four kinds of mice were used: Balb/c, CBA, C57Bl/6 and outbred white laboratory mice. Groups composed of tens mice each were studied as control and as experimental cohorts. Mice were treated with RSHG on alternate days throughout the experiment. Balsam (2 drops ≈ 100 μl) was applied on the back between the shoulders and spread. In the process of experiments, the coats of all animals looked well and we could not distinguish between experimental and control animals. Also we did not observe any behavioral peculiarities of experimental mice.

As RSHG is designed to stimulate hair growth, we could suppose that it should increase the hair follicle sizes, so we measured them. For this purpose we searched maximal size follicles on the photos of histological sections. Only one follicle per mouse was included. We sorted these follicles in a descending order (fig.2-3).

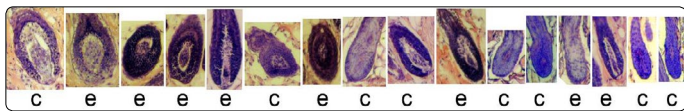


Figure 2. RSHG influence on hair follicles size

This row corresponds to experiment №1 from fig.2. The follicles are sorted in a descending order. c control. e experiment

mice	sex	duration of experiments (months)	Maximal follicles diameter sorted in a descending order within each experiment	exp. №
white laboratory mice	♂	4	c e e e e c c c c c c c c c c	1
	♀	4	e e c c c c c c c c c c c c	2
CBA	♂	6	e e c c c e e c c c c c c c c	3
	♀	24	c e e e c c c c c c c c c c c	4
Balb/c	♂	6	e e c c c c c c c c c c c e c	5
	♀	5	e e c c c c c c c c c c c c c	6
Balb/c	♀	5	e c c c c c c c c c c c c c c	7
	♀	24	e e c c c c c c c c c c c c c	8
C57Black/6	♂	12	e c c e e c c e e c c c c c c	9
	♀	12	e e c c e e c c c c c c c c c	10
C57Black/6	♀	24	c e e e c c e c c c c c c c c	11

Figure 3. RSHG influence on hair follicles size. e - experimental, c - control.

As one can see, there is a tendency of follicle size increase in each of eleven experiments. Experimental follicles are located more left-of-center in the rows than controls. As it occurs in eleven independent experiments, the probability of a mistake is less than 0,001 (1/2¹¹).

Long term RSHG application increases volumes of hair roots

To have additional evidence of the effect of RSHG on follicle size we performed measurements of the volumes of plucked hair roots (fig. 4a). We measured roots from 5 control and 5 experimental mice (25-30 hairs from each) and made ascending distributions (fig 4c and 4d).

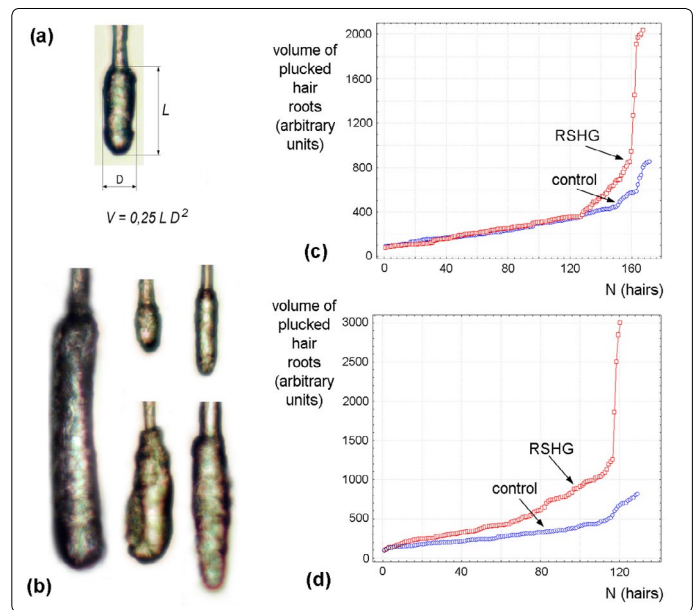


Figure 4. Influence of RSHG on hair roots volume distribution

(a) Scheme of root volume calculation, (b) - examples of various roots, (c) results of 130 days RSHG application to females of outbred white laboratory mice, (d) results of 90 days of RSHG application to CBA males.

Figures 4c and 4d show the significant (p < 0,01) increase of hair roots volumes after RSHG application. At the same time, it is clear that volumes of the smallest hair roots are not affected by RSHG. Instead, large hair root are sensitive to RSHG. Especially responsive are a few percent of hairs with the largest root volumes, which increased 2.5 - 3.75 times.

Long term RSHG application increases skin thickness

The increase of the mean follicle size without affecting the sizes of smallest follicles can indicate that RSHG increases the portion of anagen hairs. So far as in mice there are growth waves of hairs which are matched with the increasing of skin thickness [2] we compare the skin thickness as well.

Fig. 5 represents the results of these experiments. They are very similar to the follicle sizes results. Each of ten experiments showed a tendency to the increase of skin thickness under the influence of RSHG. The probability of a mistake was less than 0,001 (1/2¹⁰).

mice	sex	duration of experiments (months)	Skin thickness sorted in a descending order within each experiment	exp. №
white laboratory mice	♂	4	e e e c c c c e e c c c c c c c	1
	♀	4	e e c c c c c c c c c c c c	2
CBA	♂	6	e e c c e e c c e e c c c c c c	3
	♀	24	e e c c e e c c c c c c c c c c	4
CBA	♀	6	c e e c c e e c e e c c c c c c	5
	♂	5	e e e c c c c c c e e c c c c	6
Balb/c	♀	5	e c c c c e e e c c c c c c c	7
	♀	24	e e c c c c c c c c c c c c c c	8
C57Black/6	♂	12	e e c c e e c c c c c c c c c c	9
	♀	12	e c c e e c e e e e c c c c c c	10

Figure 5. RSHG influence on skin thickness. e - experimental, c - control.

Some of the results are presented in fig. 6. Histological analysis was performed after different periods of RSHG applications (from 4 to 24 months). We found that RSHG treatment did not abolish the waves of hair growth (fig 6a). Alterations of skin thickness were mainly due to the variability of the thickness of adipose.

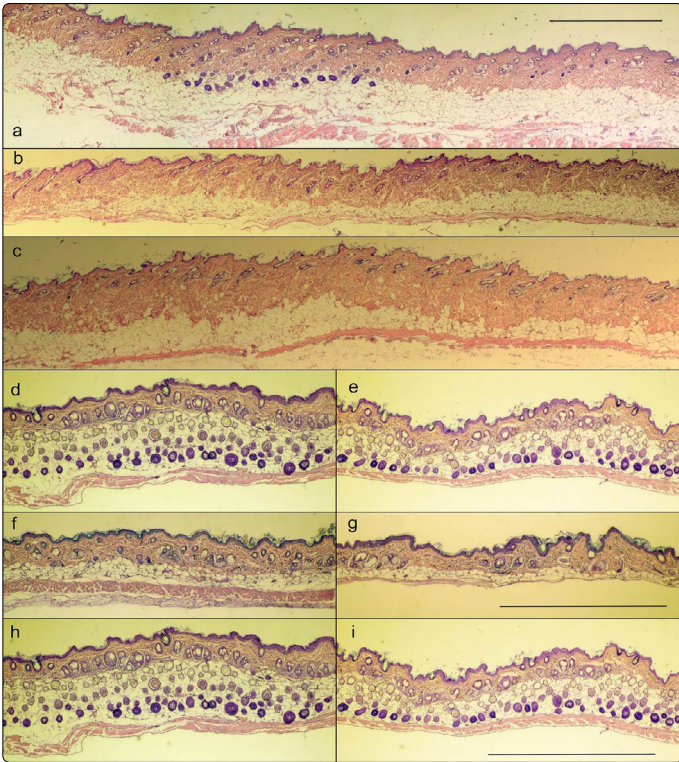


Figure 6. RSHG influence on mice skin thickness (a-c) CBA males, the rest - Balb/c females. (a) and (c) - 6 months RSHG application, (b) - control. RSHG application does not abolish hair growth waves (a). (d) and (f) - 12 months RSHG application, (e) and (g) - control. (h) - 24 months RSHG application, (i) - control. Bars 1 mm. Groups of photos (a-c), (d-g) and (h-i) have the same magnifications. Hematoxylin-eosin staining, digital contrast.

RSHG application stimulates overgrowth of bald areas of old mice skin

We wanted to find a clear manifestation of RSHG action in animals. Young mice have good hair, so it is difficult to strongly stimulate their growth. We decided to work with old black mice, which exhibit clear defects of hair growth. In a considerable population (around one hundred) of more than two years old C57Black/6 mice we choose two groups of five mice with visible alopecia areas. After two months of RSHG application, alopecia regions were overgrown by hair in both groups (fig. 7).

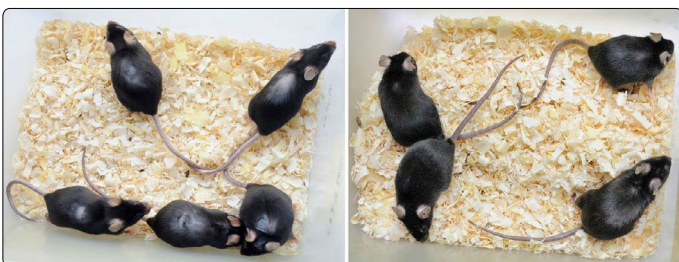


Figure 7. Decrease of bald areas on the skin of senescent C57Black/6 mice after 2 months of RSHG application. Left - before application, right - after application.

Transcriptome analysis is a modern investigation technique that permits to entirely cover alterations of cellular gene expression. We chose to use it in the process of RSHG investigation because due to the complex composition RSHG can simultaneously act in different directions.

At present, we do not know that cells are the target of RSHG *in vivo*. Those may be hair keratinocytes, dermal papilla cells, dermal fibroblasts, cells of immune system, nerve cells etc. Selective and rapid isolation of these cells from murine tissues without changing their pattern of gene expression is at present impossible.

Thus, we have selected as a model the culture of adult human skin fibroblasts. This culture (strain 1608) was comprehensively investigated [24]. Proliferative capabilities of cells allow working with a homogeneous cell population where 90% cells are in cell cycle.

The transcriptome analysis using the MetaCore program showed with a high reliability that among most significant processes enriched in differentially expressed genes (Pathway Maps) was cell cycle, cytoskeleton dynamics, epithelial-to-mesenchymal transition (EMT), cell adhesion and extracellular matrix remodeling (fig. 8).

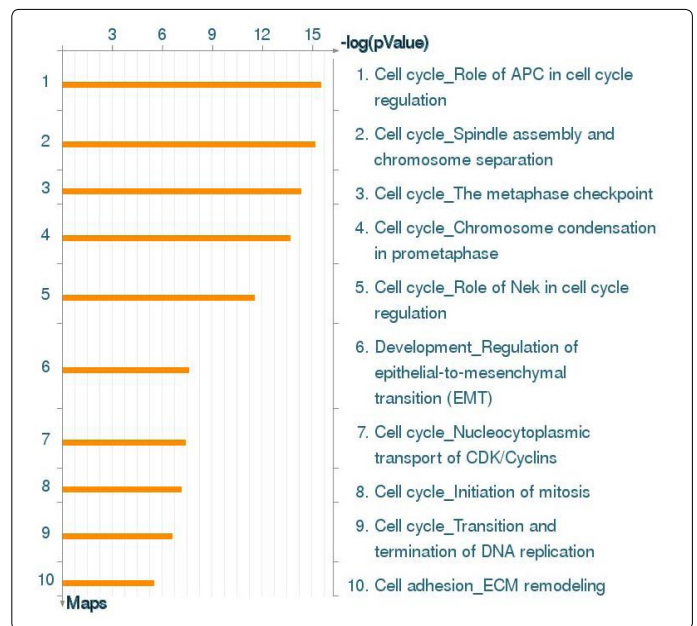


Figure 8. Pathway Maps under the influence of RSHG.

Networks analysis (ProcessNetworks) revealed that most significant changes are observed in cytoskeleton (-log (pValue)=23), cell cycle (-log (pValue)=22), cell adhesion and cell matrix (-log (pValue)=8).

Metabolic Networks analysis revealed activation of the N-acyl-sphingosine phosphate and sphingomyelin pathways with [-log (pValue) = 7], (fig. 9).

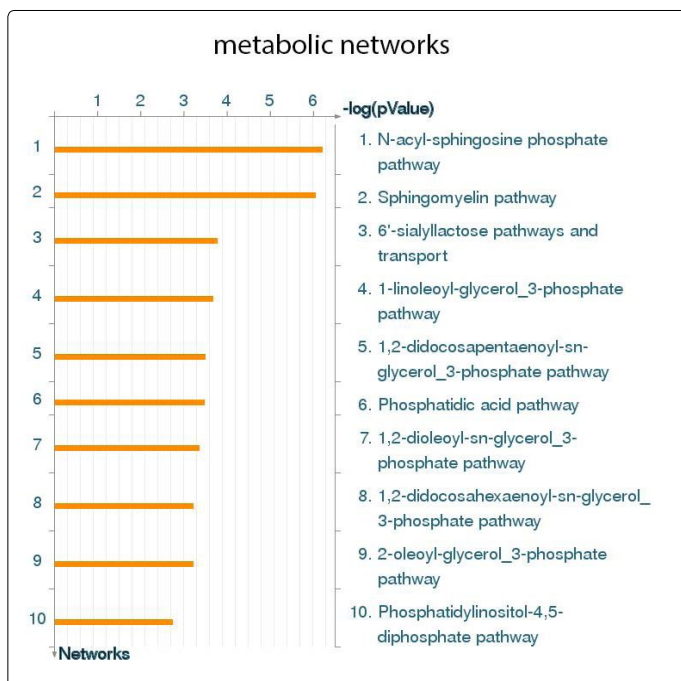


Figure 9. Changes in metabolic networks under the influence of RSHG.

Gene Ontology Processes analysis pointed again to the cell cycle and various developmental processes.

Discussion

During the primary study, we found that biggest follicles were observed in RSHG treated specimens. In the course of quantitative analysis, we confirmed that indeed RSHG treatment results in the increase of follicle size (fig. 2-3). There are several types of hairs on the back of the mouse. A few percents of the largest hairs constitute guard hairs. These hairs are also known as tylotrich hairs and they exhibit particular mechanisms and timing of embryonic development. Unlike other, hairs they have a connection with the nervous system to perform a sensory function [25]. On histological sections, guard hair follicles can be identified by their exceptional size. Probably the RSHG affects guard hairs especially strongly.

The second approach was the investigation of plucked hair roots. The plucked hair shaft has been used in medical research over the last 60 years [26]. Gho and colleagues demonstrated that most of the epithelial structures from the hair follicle remain attached to the plucked hair [27]. The maintenance of integrity of the outer root sheet after hair plucking has been documented by Limat and colleagues [28]. The outer root sheet is visible as a white layer covering the root [29]. Hair follicle stem cells lie in the bulge of the outer root sheet and give rise to keratinocytes and other cell types.

Because the size of the follicle is determined by cell number, we chose the hair root volume as the best parameter to characterize the follicle size. To simplify the task, we calculated the volumes circumscribed around root cylinders. Results obtained with this approach were similar to the results obtained by studying histological sections but much more reliable. Moreover this approach provided hair roots volumes distribution that showed that RSHG did not change the sizes

of the smallest (resting) roots (fig. 4c and d), but increased the sizes of the biggest ones.

Mice hair coat contains a mix of four different types of hair: guard, awl, auchene and zigzag, as well as specialized hairs such as vibrissae and tail hair [25]. Guard hairs are the largest and constitute 1-3% [25]. As it can be seen from fig. 4, the experimental curves have sharp bends that separate 4.7% (fig. 4c) and 4.2% (fig. 4d) of the biggest hairs. Probably the most prominent action of RSHG is directed to guard hairs.

Although the maximal effect was observed in guard hairs, RSHG action was not restricted to them. As it can be seen from fig. 4, a weaker action can overlap about 25% of the hairs in case of fig. 4c and most of the hairs in fig 4d. This observation is consistent with the suggestion that RSHG increases the ratio of anagen hairs.

Baldness development is typically due to follicles miniaturization. So the follicles enlargement under the influence of RSHG can be considered as an anti-balding action.

Well-developed adipose is a necessary precondition for hair growth. At the beginning of the growth, hair follicles increase their volumes through cell multiplication and immerse deeply into adipose. In animals with coordinated hair replacement the overall skin thickness increases in response to massive anagen initiation at the expense of subcutis elaboration [2]. There are naturally determined waves of hair growth during mice development. The ordering of these waves is very regular initially after birth. With passing time they breakdown into "islands of growth", especially on the dorsum [30]. First, we observed that RSHG application does not abolish hair growth waves. Second, we found the increase of skin thickness (Fig. 6), and also detected a significant alteration of subcutis after RSHG treatment (fig. 6).

Previously we showed that RSHG possesses similar activity in humans [21]. We revealed substantial restoration of hair growth, augmentation of subcutis and an overall skin structure normalization on the basis of histological examination of scalps of two volunteers after course of RSHG treatment. It has also been shown that RSHG induces adipogenic differentiation of mesenchymal stromal cells in culture [21]. Based on these results, we suggest that RSHG stimulates adipose elaboration.

Increases of follicle size and of skin thickness are indirect signs of hair growth stimulation. Experiments with aging mice showed that RSHG indeed stimulates hair growth.

To understand the possible mechanisms of RSHG action, we performed transcriptome studies. Analysis reveals some alterations of cell cycle, processes of development, stimulation of epithelial-to-mesenchymal transition (EMT), extracellular matrix (ECM) remodeling and lipid metabolism pathways. Considering that the RSHG mainly consists of lipid derivatives (Fig. 1), it is clear that it must change the metabolism of lipids. Activation of EMT in case of fibroblasts means increase of motility and cytoskeleton alterations. ECM remodeling and changes of cells adhesion are also connected to EMT.

Observed alterations of cell cycle gene expression suggest that RSHG action leads to accumulation of cells in the G2 period.

Such accumulation is typical for keratinocytes [31]. This result was not unexpected, because previously we have shown that RSHG does not stimulate proliferation of various cells in vitro [22].

Obtained results together with earlier ones allow us to propose hypothesis about mechanisms of RSHG action. Previously we demonstrated that RSHG increases cell death under conditions of oxidative stress [22]. Oxidative stress is known to induce cell damage and senescence in vivo. Selective elimination of senescent cells in the body decreases senescence associated pathology [32]. Probably the same situation takes place in hair follicles: decreasing senescent cells ratio has an anti-aging effect.

After application to the skin, RSHG needs to overcome the layer of cornified keratinocytes to realize its action. Another way is transfollicular transport [33]. There is a gap around hair and any preparation including nanoparticles [34] can easily achieve follicular region. Thus, maximal delivery of RSHG should be addressed to follicle cells. If there are senescent or damaged cells within the follicle, RSHG could induce their elimination that should stimulate regeneration process and hair growth.

Apparently the main locale of cell aging in skin is connective tissue including dermal papilla and follicles envelop. Indeed, connective tissue develops aging markers more rapidly than epithelial tissue [35]. Dermal papilla is the principal regulator of hair growth [36]. Moreover, it was shown that hair stem cells (epithelial cells) are stored in the skin even after fully developed baldness [37].

During aging, the decrease of stem cell functions is determined not only by alteration of stem cells *per se* but also by aging of stem cell niche. Thus, the function of hematopoietic stem cells depends on stromal cells of bone marrow [38]; the function of myogenic stem cells depends on myogenic niche [39].

Considering a hair follicle as a system composed of two components, connective tissue and epithelium, one can notice a principal difference of their lifetime. Epithelial cells actively proliferate during each hair cycle and the new groups of stem cells go from the bulge to maintain new rounds of proliferation. At the same time, dermal papilla growth is restricted and their cells should develop senescent phenotype relatively rapidly as any slowly dividing or non-dividing cell.

We suggest that RSHG action is not restricted to the stimulation of senescent cells elimination. Previously we have shown that RSHG induce autophagy [22]. Mild autophagy exhibits anti-aging activity [40]. It cleans cells from damaged proteins and organelles, hastens their turnover and therefore improves cells functions. Autophagy induction is capable to preserve stem cells and inhibit the development of baldness in mice with increased expression of Wnt1 [41].

On the other hand, autophagy induction leads to production of high energy compounds that hasten cell metabolism, as it happens with autophagy induction in tumor-associated fibroblasts [42]. Such processes can support both hair growth and subcutis elaboration, a precondition of anagen onset. However, excessive autophagy can increase cell

death [43], and, probably, senescent and damaged cells are more sensitive to it than healthy cells.

Conclusions

We suggest that a possible mechanism of action of RSHG can be selective elimination of senescent cells in hair follicles. Such preparation as RSHG should stimulate the hair growth in various kind of alopecia. The molecular mechanisms of RSHG action require further investigation.

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Conflict of Interest

No conflicts of interest have been declared.

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